

Synthesis and immunosuppressive activity evaluation of substituted *N*-imidazolidin-2-ones and *N*-tetrahydropyrimidin-2(1*H*)-ones

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Abstract

Seventeen compounds with either an imidazolin-2-one or a tetrahydropyrimidin-2(1*H*)-one scaffold were synthesized and evaluated for their immunosuppressive activity in a concanavallin A (ConA)-stimulated mouse splenocytes proliferation test. Three of these molecules exerted a significant activity at 90 μ M. All the compounds of the tetrahydropyrimidin-2(1*H*)-one series have turned out to be inactive showing the crucial role of the imidazolidin-2-one scaffold in the induction of an immunosuppressive activity.

Keywords: *Imidazolidin-2-one, tetrahydropyrimidin-2(1H)-one, urea, immunosuppressant agent*

Introduction

The control of pathological or deleterious immune responses is very often achieved by an immunosuppressive therapy. Immunosuppressant drugs are mainly used in organ transplantation for the prevention and the treatment of allograft rejection. At the present time, these molecules are also part of autoimmune diseases therapy. Some of these agents can, for example, be used in type I diabetes mellitus [1], arthritis [2,3] and dermatological pathology like psoriasis [4] or systemic lupus erythematosus [5]. In absence of immunosuppression, transplanted organs invariably undergo progressive immune-mediated injury. Acute allograft rejection is primarily mediated by immunological mechanisms implying the activation of T lymphocytes by antigen-presenting cells (APCs). Indeed, recipient T cells have the ability to recognize, through their antigen receptor, donor alloantigens presented by APCs. Once activated, T-cells differentiate, proliferate and become able to damage graft target tissues. T cells also secrete cytokines that

directly cause tissue destruction (e.g., tumor necrosis factor- β) or recruit and activate cells of the innate immune system (e.g., macrophages), which participate to the graft rejection. Current immunosuppressive agents inhibit T-cell responses either directly or through actions on APCs. These drugs can be classified to five groups in regard to their mechanism of action: inhibitors of cytokine production (calcineurin inhibitors such as cyclosporine and tacrolimus), inhibitors of cytokine binding (IL-2 receptor α chain specific monoclonal antibody), inhibitors of cytokine receptor signal transduction (rapamycin), inhibitors of DNA synthesis (cyclophosphamide, azathioprine, mycophenolate mofetil, leflunomide, brequinar sodium, methotrexate) and inhibitors of APC development and maturation (glucocorticoids, rapamycin). Although these agents, over the past 40 years, have transformed solid organ transplantation into a routine clinical procedure with a satisfactory control of acute rejection and adequate short-term graft survival [6], several problems remain. First, these drugs exhibit important side effects due to their

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intrinsic toxicity (e.g., nephrotoxicity of calcineurin inhibitors, hematotoxicity of mycophenolate mofetil, myelotoxicity of rapamycin, etc.) and to their lack of specificity, which triggers a general immunodepression state responsible for an enhanced risk of opportunistic infections and neoplastic complications [7]. These adverse effects often compromise patient and graft survival. Moreover these drugs have low efficiency on chronic graft rejection, which is often responsible for long-term graft loss [8].

In a previous work, we described the synthesis and SAR of a series of imidazolidin-2-ones, which exhibit immunosuppressive properties [9]. These studies permitted us to identify a lead compound **1** (Figure 1), which has shown maximal inhibition of the mouse splenocytes Con-A-induced proliferation at 30 μ M. These results are comparable to those obtained with the positive control, cyclosporine A, at 5 μ M (optimal dose). However, this molecule exerts cytotoxic effects on human MRC5 fibroblasts used in our cytotoxicity assay with an IC₅₀ of 21 μ M and, so, an unsatisfactory toxicity/activity index of 0.7. These interesting results prompted us to synthesize some derivatives of **1**. To explore the role of the imidazolidin-2-one scaffold in the emergence of immunosuppressive activity, we first decided to expand the ureic cycle by preparing some tetrahydropyrimidin-2-(1*H*)-one derivatives. We then synthesized some analogues of **1** with an imidazolidin-2-one moiety *N*-substituted by a phenyl or azaheterocyclic groups like phthalimidic moieties by analogy with some thalidomide analogues which exert TNF- α production inhibitory properties.

Materials and methods

General

Melting points were determined on a Tottoli-Büchi apparatus (Büchi, Flawil, Switzerland) and are uncorrected. Structures of the described compounds were supported by IR, ¹H-NMR and microanalytical data. IR spectra were run with KBr pellets on a Perkin-Elmer FT-IR Paragon 1000 grating infrared spectrometer (Perkin-Elmer, St-Quentin-en-Yvelines, France). ¹H-NMR spectra were recorded on a Bruker AC 250 spectrometer (250 MHz) (Bruker, Wissembourg, France), using CDCl₃ or DMSO as a solvent;

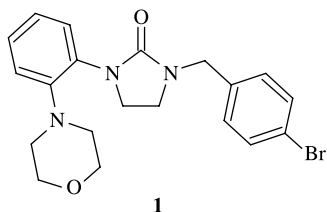


Figure 1. Structure of compound **1**.

chemical shifts (δ) are reported in parts per million (ppm), from internal Me₄Si. Mass spectra were recorded on an ESQUIRE-LC spectrometer (Bruker) (electrospray ionisation with ion trap system). Purification of synthesized compounds was made using columns of silica gel (Silica gel 60, 70-230 mesh, E. Merck, Darmstadt, Germany), with appropriate solvents. Anhydrous Na₂SO₄ was always used as the drying agent. Chemicals were purchased from Sigma-Aldrich Fluka (St Quentin Fallavier, France), Lancaster Synthesis (Bisheim, France) or Avocado (La Tour du Pin, France).

Chemistry

The synthesis of *N*-substituted imidazolidin-2-ones **10-17** and **33-37**, from ureas **2-9** and **28-32** respectively, and of *N*-substituted tétrahydropyrimidin-2(1*H*)-ones **42-45**, from ureas **38-41**, is shown in Scheme 1.

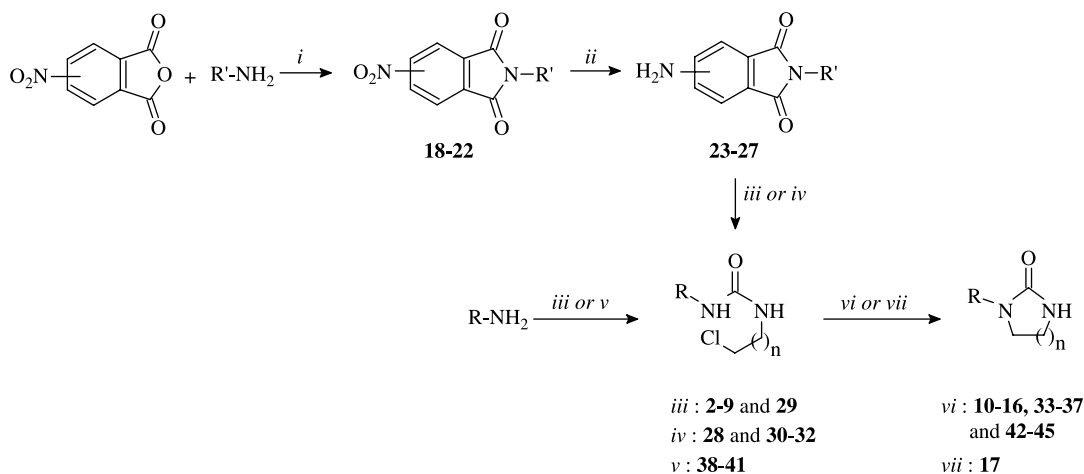
1-(4-Bromophenyl)-3-(2-chloroethyl)urea (2). To a solution of 4-bromoaniline (3 g, 17.40 mmol) in chloroform (50 mL) was added 2-chloroethyl isocyanate (1.51 mL, 17.40 mmol). The mixture was refluxed for 40 min, and then the solvent was removed under reduced pressure. The crystalline residue was recrystallized from diethyl ether to give compound **2** as a white powder. M.p. = 177°C, Yield = 96%. IR (KBr) (ν , cm⁻¹) 3317 (NH), 1631 (C=O), 825 (C-Cl), 1071 (C-Br). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.40-3.52 (m, 2H, CH₂N), 3.69 (t, 2H, CH₂Cl, ³J = 5.8), 6.48 (t, 1H, NH³, ³J' = 5.8), 8.84 (s, 1H, NH¹), 7.30-7.50 (m, 4H, H_{arom}).

Ureas **3-9** and compound **29** were also synthesized according to this procedure with a reflux time in a range of 5 min to 39 h.

1-(2-Chloroethyl)-3-(3-chlorophenyl)urea (3). Recrystallized from diethyl ether. M.p. = 99°C, Yield = 65%. IR (KBr) (ν , cm⁻¹) 3357 (NH), 1638 (C=O), 1076 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 3.55-3.64 (m, 4H, CH₂-CH₂Cl), 5.89 (bs, 1H, NH³), 7.52 (s, 1H, NH¹), 6.97-7.02 (m, 1H, H⁴), 7.14-7.17 (m, 2H, H^{5'} and H^{6'}), 7.34 (dd, 1H, H^{2'}, ⁴J = ⁴J' = 1.8).

1-(2-Chloroethyl)-3-(4-methylthiophenyl)urea (4). Recrystallized from chloroform. M.p. = 123°C, Yield = 48%. IR (KBr) (ν , cm⁻¹) 3334 (NH), 1636 (C=O), 818 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 2.47 (s, 3H, CH₃), 3.56-3.68 (m, 4H, CH₂-CH₂Cl), 5.53 (bs, 1H, NH³), 6.95 (s, 1H, NH¹), 7.21-7.27 (m, 4H, H_{arom}).

1-(2-Chloroethyl)-3-(2-phenoxyphenyl)urea (5). Recrystallized from diethyl ether. M.p. = 130°C, Yield = 74%. IR (KBr) (ν , cm⁻¹) 3341 (NH), 1641



Scheme 1. Synthesis of imidazolidin-2-ones 10-17 and 33-37 and tetrahydropyrimidin-2(1H)-ones 42-45. Reaction reagents and conditions: (i) AcOH, reflux; (ii) Pd/C 5%, H₂, THF, 50°C; (iii) 2-chloroethyl isocyanate, CHCl₃, reflux; (iv) 2-chloroethyl isocyanate (8 eq), microwaves, 82°C, 20 W; (v) 3-chloropropyl isocyanate, CHCl₃, reflux; (vi) Cs₂CO₃, CH₃CN, reflux; (vii) Na₂CO₃, CH₃CN, reflux.

(C=O), 750 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ, ppm) 3.55-3.65 (m, 4H, CH₂-CH₂Cl), 5.40 (bs, 1H, NH³), 6.84 (dd, 1H, H^{3'}, ³J = 8.0, ⁴J = 1.6), 6.94 (dd, 1H, H^{4'}, ³J = ³J' = 8.0), 6.98 (d, 2H, H^{2''} and H^{6''}, ³J''' = 8.0), 7.03 (s, 1H, NH¹), 7.08-7.14 (m, 2H, H^{5'} and H^{4''}), 7.33 (dd, 2H, H^{3''} and H^{5''}, ³J''' = ³J''' = 8.0), 8.11 (dd, 1H, H^{6'}, ³J'' = 8.0, ⁴J = 1.2).

1-(2-Chloroethyl)-3-(4-chloro-3-trifluoromethylphenyl)urea (6). Recrystallized from diisopropyl ether. M.p. = 123°C, Yield = 50%. IR (KBr) (ν, cm⁻¹) 3366 (NH), 1654 (C=O), 1029 and 829 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ, ppm) 3.52-3.72 (m, 4H, CH₂-CH₂Cl), 5.96 (bs, 1H, NH³), 7.81 (s, 1H, NH¹), 7.29-7.41 (m, 2H, H^{5'} and H^{6'}), 7.58 (d, 1H, H^{2'}, ⁴J = 2.1).

1-(2-Chloroethyl)-3-(1,3-dimethyl(1H)pyrazol-5-yl)urea (7). Recrystallized from dichloromethane/diethyl ether (50/50). M.p. = 128°C, Yield = 35%. IR (KBr) (ν, cm⁻¹) 3327 (NH), 1687 (C=O), 782 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ, ppm) 2.25 (s, 3H, CH₃), 3.51-3.63 (m, 2H, CH₂N), 3.64 (t, 2H, CH₂Cl, ³J = 5.8), 3.71 (s, 3H, NCH₃), 5.30 (bs, 1H, NH³), 5.97 (s, 1H, H_{pyraz}), 6.66 (s, 1H, NH¹).

1-(2-Chloroethyl)-3-(quinolin-8-yl)urea (8). Recrystallized from diethyl ether. M.p. = 157°C, Yield = 84%. IR (KBr) (ν, cm⁻¹) 3309 (NH), 1648 (C=O), 823 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ, ppm) 3.68-3.78 (m, 4H, CH₂-CH₂Cl), 5.61 (bs, 1H, NH³), 7.39-7.45 (m, 2H, H^{3'} and H^{5'}), 7.52 (dd, 1H, H^{6'}, ³J = ³J' = 7.6), 8.15 (dd, 1H, H^{4'}, ³J'' = 8.3, ⁴J = 1.6), 8.53 (dd, 1H, H^{7'}, ³J = 7.6, ⁴J = 1.2), 8.74 (dd, 1H, H^{2'}, ³J''' = 4.2, ⁴J = 1.6), 9.10 (s, 1H, NH¹).

1-(2-Chloroethyl)-3-(1H-indol-5-yl)urea (9). Recrystallized from diethyl ether. M.p. = 153°C, Yield = 90%. IR (KBr) (ν, cm⁻¹) 3421 (NH_{indol}),

3315 (NH), 1626 (C=O), 734 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ, ppm) 3.39-3.51 (m, 2H, CH₂N), 3.69 (t, 2H, CH₂Cl, ³J = 5.8), 6.24 (s, 1H, H^{3'}), 6.30 (t, 1H, NH³, ³J' = 5.8), 7.14-7.18 (m, 1H, H^{6'}), 7.24-7.28 (m, 2H, H^{2'} and H^{7'}), 7.87 (s, 1H, H^{4'}), 8.37 (s, 1H, NH¹), 10.92 (s, 1H, NH_{indol}).

1-(2-Chloroethyl)-3-(2-morpholin-4-yl-1,3-dioxo-2,3-dihydro-1H-isoindol-5-yl)urea (29). Recrystallized from diethyl ether. M.p. = 207°C, Yield = 70%. IR (KBr) (ν, cm⁻¹) 3354 (NH), 1773 and 1716 (C=Oimide), 1653 (C=O), 738 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ, ppm) 3.33-3.38 (m, 4H, CH₂N_{morphol}), 3.44-3.61 (m, 2H, CH₂N), 3.70-3.81 (m, 6H, CH₂Cl and CH₂O), 6.74 (bs, 1H, NH³), 7.68 (d, 1H, H^{6'}, ³J = 8.2), 7.81 (d, 1H, H^{7'}, ³J = 8.2), 8.07 (s, 1H, H^{4'}), 9.49 (s, 1H, NH¹).

1-(2-Chloroethyl)-3-(2-morpholin-4-yl-1,3-dioxo-2,3-dihydro-1H-isoindol-4-yl)urea (28). In a sealed tube containing 3-aminophthalimide (0.51 g, 2.06 mmol) was added 2-chloroethyl isocyanate (1.43 mL, 16.48 mmol). The mixture was stirred and heated by microwaves at 82°C with a 20 W power during 40 min and taken up into acetone. The solvent was then evaporated under reduced pressure and purification was accomplished by column chromatography over silica gel with dichloromethane. The residue was recrystallized from diethyl ether to give urea 28 as a white powder. M.p. = 226°C, Yield = 79%. IR (KBr) (ν, cm⁻¹) 3328 (NH), 1772 and 1712 (C=Oimide), 1700 (C=O), 745 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ, ppm) 3.33-3.38 (m, 4H, CH₂N_{morphol}), 3.42-3.54 (m, 2H, CH₂N), 3.60-3.85 (m, 6H, CH₂O and CH₂Cl), 7.39 (d, 1H, H^{7'}, ³J = 7.0), 7.73 (dd, 1H, H^{6'}, ³J = 7.0, ³J' = 8.5), 8.13

(bs, 1H, NH³), 8.57 (d, 1H, H^{5'}, ³J' = 8.5), 8.94 (s, 1H, NH¹).

Ureas **30-32** were prepared according to the same procedure.

1-(2-Chloroethyl)-3-(2-phenyl-1,3-dioxo-2,3-dihydro-1H-isoindol-4-yl)urea (30). Recrystallized from diethyl ether. M.p. = 193°C, Yield = 66%. IR (KBr) (ν , cm⁻¹) 3391 (NH), 1753 and 1700 (C=Oimide), 1682 (C=O), 767 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.43-3.55 (m, 2H, CH₂N), 3.73 (t, 2H, CH₂Cl, ³J = 5.8), 7.40-7.80 (m, 6H, H_{arom} and H^{7'}), 7.80 (dd, 1H, H^{6'}, ³J' = 7.3, ³J'' = 8.5), 8.16 (t, 1H, NH³, ³J = 5.8), 8.60 (d, 1H, H^{5'}, ³J'' = 8.5), 9.04 (s, 1H, NH¹).

1-(2-Chloroethyl)-3-(2-phenyl-1,3-dioxo-2,3-dihydro-1H-isoindol-5-yl)urea (31). Recrystallized from diethyl ether. M.p. = 230°C, Yield = 75%. IR (KBr) (ν , cm⁻¹) 3359 (NH), 1774 and 1717 (C=Oimide), 1703 (C=O), 748 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.48-3.57 (m, 2H, CH₂N), 3.74 (t, 2H, CH₂Cl, ³J = 5.8), 6.77 (t, 1H, NH³, ³J = 5.8), 7.43-7.59 (m, 5H, H_{arom}), 7.71 (dd, 1H, H^{6'}, ³J' = 8.2, ⁴J = 1.8), 7.86 (d, 1H, H^{7'}, ³J' = 8.2), 8.19 (d, 1H, H^{4'}, ⁴J = 1.8), 9.55 (s, 1H, NH¹).

1-(2-Chloroethyl)-3-(2-benzyl-1,3-dioxo-2,3-dihydro-1H-isoindol-4-yl)urea (32). Recrystallized from dichloromethane. M.p. = 186°C, Yield = 41%. IR (KBr) (ν , cm⁻¹) 3317 (NH), 1760 and 1708 (C=Oimide), 1659 (C=O), 740 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.42-3.54 (m, 2H, CH₂N), 3.71 (t, 2H, CH₂Cl, ³J = 5.2), 4.78 (s, 2H, CH₂Ph), 7.20-7.40 (m, 5H, H_{arom}), 7.44 (d, 1H, H^{7'}, ³J' = 6.7), 7.73 (dd, 1H, H^{6'}, ³J' = 6.7, ³J'' = 8.5), 8.11 (t, 1H, NH³, ³J = 5.2), 8.58 (d, 1H, H^{5'}, ³J'' = 8.5), 8.97 (s, 1H, NH¹).

1-(4-Chlorophenyl)-3-(3-chloropropyl)urea (38). To a solution of 4-chloroaniline (1 g, 7.84 mmol) in chloroform (50 mL) was added dropwise 3-chloropropyl isocyanate (0.81 mL, 7.84 mmol). The mixture was refluxed for 1 h, and then the solvent was removed under reduced pressure. The crystalline residue was recrystallized from diethylether to give compound **38** as a white powder. M.p. = 143°C, Yield = 95%. IR (KBr) (ν , cm⁻¹) 3331 (NH), 1635 (C=O), 828 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 1.86-1.98 (m, 2H, CH₂CH₂CH₂), 3.18-3.30 (m, 2H, CH₂NH), 3.70 (t, 2H, CH₂Cl, ³J = 6.6), 6.33 (t, 1H, NH³, ³J' = 6.6), 7.29 (d, 2H, H^{3'} and H^{5'}, ³J'' = 8.9), 7.45 (d, 1H, H^{2'} and H^{6'}, ³J'' = 8.9), 8.63 (s, 1H, NH¹).

Ureas **39-41** were synthesized according to this method.

1-(3-Chloro-4-fluorophenyl)-3-(3-chloropropyl)-urea (39). Recrystallized from diethyl ether.

M.p. = 107°C, Yield = 84%. IR (KBr) (ν , cm⁻¹) 3336 (NH), 1651 (C=O), 1217 (C-F), 1052 and 797 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 1.87-1.99 (m, 2H, CH₂CH₂CH₂), 3.29-3.41 (m, 2H, CH₂NH), 3.56 (t, 2H, CH₂Cl, ³J = 6.2), 6.02 (t, 1H, NH³, ³J' = 6.2), 6.91-7.00 (m, 2H, H^{5'} and H^{6'}), 7.37 (dd, 1H, H^{2'}, ⁴J_{HF} = 6.5, ⁴J = 2.4), 7.95 (s, 1H, NH¹).

1-(3-Chloropropyl)-3-(2-methoxy-5-trifluoromethylphenyl)urea (40). Recrystallized from diethyl ether. M.p. = 158°C, Yield = 68%. IR (KBr) (ν , cm⁻¹) 3370 (NH), 1653 (C=O), 810 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 1.90-2.15 (m, 2H, CH₂CH₂CH₂), 3.45-3.65 (m, 4H, CH₂CH₂CH₂), 3.91 (s, 3H, OCH₃), 5.01 (bs, 1H, NH³), 6.89 (d, 1H, H^{2'}, ³J = 7.0), 6.99 (s, 1H, H^{6'}), 7.25 (d, 1H, H^{4'}, ³J = 7.0), 8.47 (s, 1H, NH¹).

1-(3-Chloro-4-cyanophenyl)-3-(3-chloropropyl)urea (41). Recrystallized from diethyl ether. M.p. = 111°C, Yield = 89%. IR (KBr) (ν , cm⁻¹) 3321 (NH), 2226 (C≡N), 1677 (C=O), 828 and 1045 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 1.80-2.00 (m, 2H, CH₂CH₂CH₂), 3.20-3.30 (m, 2H, CH₂NH), 3.65-3.75 (m, 2H, CH₂Cl), 6.65 (bs, 1H, NH³), 7.40 (d, 1H, H^{6'}, ³J = 8.8), 7.79 (d, 1H, H^{5'}, ³J = 8.8), 7.94 (s, 1H, H^{2'}).

1-(4-Bromophenyl)imidazolidin-2-one (10). Urea **2** (2 g, 7.21 mmol) was dissolved in acetonitrile (50 mL) and cesium carbonate (2.35 g, 7.21 mmol) was added. The reaction mixture was stirred and refluxed for 20 h and then filtered. The filtrate solvent was evaporated *in vacuo*. The crystalline residue was recrystallized from diethyl ether to give compound **10** as a white powder. M.p. = 185°C, Yield = 75%. MS-ES⁺ (CH₃OH) (m/z) 241 (⁸¹Br), 239 (⁷⁹Br). IR (KBr) (ν , cm⁻¹) 1681 (C=O), 1071 (C-Br). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 3.58 (t, 2H, CH₂NH, ³J = 8.2), 3.90 (t, 2H, CH₂N, ³J = 8.2), 5.53 (bs, 1H, NH), 7.40-7.45 (m, 4H, H_{arom}).

All imidazolidin-2-ones and tetrahydropyrimidin-2(1H)-ones, compound **17** excepted, were prepared according to this procedure, with a reflux time of 30 min to 19 h.

1-(3-Chlorophenyl)imidazolidin-2-one (11). Recrystallized from diethyl ether. M.p. = 121°C, Yield = 47%. MS-ES⁺ (CH₃OH) (m/z) 197 (³⁷Cl), 195 (³⁵Cl). IR (KBr) (ν , cm⁻¹) 1703 (C=O), 1080 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 3.59 (t, 2H, CH₂NH, ³J = 8.5), 3.91 (t, 2H, CH₂N, ³J = 8.5), 5.65 (bs, 1H, NH), 7.02 (dd, 1H, H^{4'}, ³J' = 8.2, ⁴J = 1.2), 7.25 (dd, 1H, H^{5'}, ³J' = ³J'' = 8.2), 7.43 (dd, 1H, H^{6'}, ³J'' = 8.2, ⁴J' = 1.2), 7.60 (dd, 1H, H^{2'}, ⁴J = ⁴J' = 1.2).

1-(4-Methylthiophenyl)imidazolidin-2-one (12). Recrystallized from diisopropyl ether. M.p. = 187°C, Yield = 32%. MS-ES⁺ (CH₃OH) (m/z) 208. IR (KBr) (ν , cm⁻¹) 1707 (C=O). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 2.47 (s, 3H, CH₃), 3.57 (t, 2H, CH₂NH, ³J = 7.3), 3.87-3.95 (m, 2H, CH₂N), 5.08 (bs, 1H, NH), 7.28 (d, 2H, H³ and H⁵, ³J' = 8.8), 7.48 (d, 2H, H² and H⁶, ³J' = 8.8).

1-(2-Phenoxyphenyl)imidazolidin-2-one (13). Recrystallized from diethyl ether. M.p. = 124°C, Yield = 48%. MS-ES⁺ (CH₃OH) (m/z) 257 ([M + 2H]⁺). IR (KBr) (ν , cm⁻¹) 1682 (C=O). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 3.42 (t, 2H, CH₂NH, ³J = 8.4), 3.89 (t, 2H, CH₂N, ³J = 8.4), 5.18 (bs, 1H, NH), 6.96-7.01 (m, 3H, H³, H^{2'} and H^{6'}), 7.09 (t, 1H, H^{4'}, ³J' = 7.5), 7.15-7.23 (m, 2H, H⁴ and H⁵), 7.32 (dd, 2H, H^{3'} and H^{5'}, ³J' = ³J'' = 7.5), 7.52 (dd, 1H, H⁶, ³J''' = 7.2, ⁴J = 2.0).

1-(4-Chloro-3-trifluoromethylphenyl)imidazolidin-2-one (14). Recrystallized from diisopropyl ether. M.p. = 148°C, Yield = 76%. MS-ES⁺ (CH₃OH) (m/z) 265. IR (KBr) (ν , cm⁻¹) 1719 (C=O), 1025 (C-Cl). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 3.55-3.70 (m, 2H, CH₂NH), 3.90-4.00 (m, 2H, CH₂N), 5.44 (bs, 1H, NH), 7.44 (d, 1H, H⁵, ³J = 8.8), 7.76 (dd, 1H, H⁶, ³J = 8.8, ⁴J = 2.8), 7.82 (d, 1H, H², ⁴J = 2.8).

1-(1,3-Dimethyl(1H)pyrazol-5-yl)imidazolidin-2-one (15). Recrystallized from diethyl ether. M.p. = 130°C, Yield = 49%. MS-ES⁺ (CH₃OH) (m/z) 180. IR (KBr) (ν , cm⁻¹) 1704 (C=O). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 2.23 (s, 3H, CH₃), 3.60 (t, 2H, CH₂NH, ³J = 7.4), 3.74 (s, 3H, NCH₃), 3.79 (t, 2H, CH₂N, ³J = 7.4), 5.50 (bs, 1H, NH), 5.88 (s, 1H, H_{pyraz}).

1-(Quinolin-8-yl)imidazolidin-2-one (16). Recrystallized from diethyl ether. M.p. = 151°C, Yield = 52%. MS-ES⁺ (CH₃OH) (m/z) 213. IR (KBr) (ν , cm⁻¹) 1702 (C=O). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.53 (t, 2H, CH₂NH, ³J = 8.3), 4.17 (t, 2H, CH₂N, ³J = 8.3), 6.81 (bs, 1H, NH), 7.60 (dd, 1H, H³, ³J' = 8.3, ³J'' = 4.1), 7.63 (dd, 1H, H⁶, ³J''' = 8.2, ³J'''' = 7.2), 7.77 (dd, 1H, H⁷, ³J'''' = 7.2, ⁴J = 1.1), 7.90 (dd, 1H, H⁵, ³J'''' = 8.2, ⁴J = 1.1), 8.44 (dd, 1H, H⁴, ³J' = 8.3, ⁴J' = 1.7), 8.95 (dd, 1H, H², ³J'' = 4.1, ⁴J' = 1.7).

1-(2-Morpholin-4-yl-1,3-dioxo-2,3-dihydro-1H-isoindol-4-yl)imidazolidin-2-one (33). Recrystallized from diethyl ether. M.p. = 193°C, Yield = 59%. MS-ES⁺ (CH₃OH) (m/z) 318 ([M + 2H]⁺). IR (KBr) (ν , cm⁻¹) 1771 and 1722 (C=Oimide), 1661 (C=O). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.25-3.35 (m, 4H, CH₂N_{morphol}), 3.49 (t, 2H, CH₂N, ³J = 7.0), 3.65-3.80 (m, 4H, CH₂O), 4.06 (t, 2H, CH₂NH,

³J = 7.0), 7.19 (bs, 1H, NH), 7.65 (d, 1H, H⁷, ³J' = 6.7), 7.78-7.88 (m, 2H, H⁵ and H⁶).

1-(2-Morpholin-4-yl-1,3-dioxo-2,3-dihydro-1H-isoindol-5-yl)imidazolidin-2-one (34). Recrystallized from diethyl ether. M.p. = 288°C, Yield = 61%. MS-ES⁺ (CH₃OH) (m/z) 318 ([M + 2H]⁺). IR (KBr) (ν , cm⁻¹) 1774 and 1719 (C=Oimide), 1655 (C=O). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.25-3.35 (m, 4H, CH₂N_{morphol}), 3.49 (t, 2H, CH₂N, ³J = 7.3), 3.65-3.80 (m, 4H, CH₂O), 4.00 (t, 2H, CH₂NH, ³J = 7.3), 7.48 (bs, 1H, NH), 7.81-7.89 (m, 2H, H⁶ and H⁷), 8.20 (s, 1H, H⁴).

1-(2-Phenyl-1,3-dioxo-2,3-dihydro-1H-isoindol-4-yl)imidazolidin-2-one (35). Recrystallized from diethyl ether. M.p. = 201°C, Yield = 87%. MS-ES⁺ (CH₃OH) (m/z) 309 ([M + 2H]⁺). IR (KBr) (ν , cm⁻¹) 1759 and 1714 (C=Oimide), 1654 (C=O). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.44-3.52 (m, 2H, CH₂N), 4.09 (t, 2H, CH₂NH, ³J = 8.1), 7.20 (bs, 1H, NH), 7.45-7.60 (m, 5H, H_{arom}), 7.77 (dd, 1H, H⁷, ³J' = 6.7, ⁴J = 1.5), 7.89 (dd, 1H, H⁶, ³J' = 6.7, ³J'' = 8.3), 7.94 (dd, 1H, H⁵, ³J'' = 8.3, ⁴J = 1.5).

1-(2-Phenyl-1,3-dioxo-2,3-dihydro-1H-isoindol-5-yl)imidazolidin-2-one (36). Recrystallized from diethyl ether. M.p. = 312°C, Yield = 78%. MS-ES⁺ (CH₃OH) (m/z) 309 ([M + 2H]⁺). IR (KBr) (ν , cm⁻¹) 1774 and 1726 (C=Oimide), 1704 (C=O). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.52 (t, 2H, CH₂N, ³J = 7.0), 4.05 (t, 2H, CH₂NH, ³J = 7.0), 7.40-7.60 (m, 6H, NH and H_{arom}), 7.90-7.95 (m, 2H, H⁶ and H⁷), 8.30 (s, 1H, H⁴).

1-(2-Benzyl-1,3-dioxo-2,3-dihydro-1H-isoindol-4-yl)imidazolidin-2-one (37). Recrystallized from ethanol. M.p. = 175°C, Yield = 39%. MS-ES⁺ (CH₃OH) (m/z) 323 ([M + 2H]⁺). IR (KBr) (ν , cm⁻¹) 1769 and 1709 (C=Oimide), 1656 (C=O). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 3.43-3.51 (m, 2H, CH₂N), 4.08 (t, 2H, CH₂NH, ³J = 7.0), 4.78 (s, 2H, CH₂), 7.21 (bs, 1H, NH), 7.30-7.40 (m, 5H, H_{arom}), 7.69 (d, 1H, H⁷, ³J' = 6.7), 7.79-7.90 (m, 2H, H⁵ and H⁶).

1-(4-Chlorophenyl)tetrahydropyrimidin-2(1H)-one (42). Recrystallized from diethyl ether. M.p. = 164°C, Yield = 86%. MS-ES⁺ (CH₃OH) (m/z) 212 (³⁷Cl), 210 (³⁵Cl). IR (KBr) (ν , cm⁻¹) 1657 (C=O), 1092 (C-Cl). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 1.93-2.01 (m, 2H, CH₂CH₂-CH₂), 3.25 (td, 2H, CH₂NH, ³J = 5.6, ⁴J = 2.5), 3.64 (t, 2H, CH₂N, ³J' = 5.6), 6.69 (bs, 1H, NH), 7.30-7.45 (m, 4H, H_{arom}).

1-(3-Chloro-4-fluorophenyl)tetrahydropyrimidin-2(1H)-one (43). Recrystallized from diethyl ether. M.p. = 153°C, Yield = 70%. MS-ES⁺ (CH₃OH) (m/z) 230 (³⁷Cl), 228 (³⁵Cl). IR (KBr) (ν , cm⁻¹)

1663 (C=O), 1223 (C-F), 1055 (C-Cl). ^1H NMR (250 MHz, CDCl_3) (δ , ppm) 2.04–2.12 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 3.41 (td, 2H, CH_2NH , $^3\text{J} = 5.8$, $^4\text{J} = 2.4$), 3.64 (t, 2H, CH_2N , $^3\text{J}' = 5.8$), 5.61 (bs, 1H, NH), 7.10 (dd, 1H, H^5 , $^3\text{J}'' = ^3\text{J}_{\text{HF}} = 8.8$), 7.19 (ddd, 1H, H^6 , $^3\text{J}'' = 8.8$, $^4\text{J}_{\text{HF}} = 4.3$, $^4\text{J} = 2.7$), 7.37 (dd, 1H, H^2 , $^4\text{J}_{\text{HF}} = 6.7$, $^4\text{J} = 2.7$).

1-(2-Methoxy-5-trifluoromethylphenyl)tetrahydropyrimidin-2(1H)-one (44). Recrystallized from diethyl ether. M.p. = 153°C, Yield = 20%. MS-ES⁺ (CH_3OH) (m/z) 276 ($[\text{M} + 2\text{H}]^+$). IR (KBr) (ν , cm^{-1}) 1665 (C=O). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 1.85–2.05 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 3.15–3.55 (m, 4H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 3.88 (s, 3H, OCH_3), 6.58 (s, 1H, NH), 7.26 (d, 1H, H^3 , $^3\text{J} = 8.6$), 7.52 (s, 1H, H^6), 7.62 (d, 1H, H^4 , $^3\text{J} = 8.6$).

1-(3-Chloro-4-cyanophenyl)tetrahydropyrimidin-2(1H)-one (45). Recrystallized from diisopropyl ether. M.p. = 169°C, Yield = 88%. MS-ES⁺ (CH_3OH) (m/z) 240 ($[\text{M} + 2\text{H}]^+$, ^{37}Cl), 238 ($[\text{M} + 2\text{H}]^+$, ^{35}Cl). IR (KBr) (ν , cm^{-1}) 2227 (C \equiv N), 1669 (C=O), 1047 (C-Cl). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 1.96–2.03 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 3.25 (td, 2H, CH_2NH , $^3\text{J} = 5.6$, $^4\text{J} = 2.4$), 3.74 (t, 2H, CH_2N , $^3\text{J}' = 5.6$), 7.12 (bs, 1H, NH), 7.54 (dd, 1H, H^6 , $^3\text{J}'' = 8.8$, $^4\text{J} = 2.0$), 7.81 (d, 1H, H^2 , $^4\text{J} = 2.0$), 7.89 (d, 1H, H^5 , $^3\text{J}'' = 8.8$).

1-(1H-indol-5-yl)imidazolidin-2-one (17). Urea **9** (0.5 g, 2.10 mmol) was dissolved in acetonitrile (50 mL) and sodium carbonate (0.22 g, 2.10 mmol) was added. The reaction mixture was stirred and refluxed for 6 h and then filtered. The filtrate solvent was removed under reduced pressure. Crystallization of the oily residue from dichloromethane/diethyl ether (50/50) gave the compound **17** as a white powder. M.p. = 256°C, Yield = 41%. MS-ES⁺ (CH_3OH) (m/z) 202 ($[\text{M} + \text{H}]^+$). IR (KBr) (ν , cm^{-1}) 3384 (NH_{indol}), 1674 (C=O). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 3.78 (t, 2H, CH_2NH , $^3\text{J} = 8.2$), 4.24 (t, 2H, CH_2N , $^3\text{J} = 8.2$), 6.34 (s, 1H, H^3), 7.16 (d, 1H, H^6 , $^3\text{J}' = 8.8$), 7.24–7.28 (m, 2H, H^2 and H^7), 7.87 (s, 1H, H^4), 8.70 (bs, 1H, NH), 10.88 (s, 1H, NH_{indol}).

2-Morpholin-4-yl-4-nitro-1H-isoindole-1,3(2H)-dione (18). To a solution of 3-nitrophthalic anhydride (2 g, 10.36 mmol) in glacial acetic acid (15 mL) was added *N*-aminomorpholine (1 mL, 10.36 mmol). The reaction mixture was stirred and refluxed for 19 h and then evaporated *in vacuo*. The crystalline residue was taken up into a solution of sodium hydrogenocarbonate (4%), filtered, washed with water, dried and recrystallized from ethanol to give **18** as a yellow powder. M.p. = 189°C, Yield = 79%.

IR (KBr) (ν , cm^{-1}) 1795 and 1729 (C=Oimide), 1530 and 1360 (NO_2). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 3.25–3.40 (m, 4H, CH_2N), 3.70–3.80 (m, 4H, CH_2O), 8.09 (dd, 1H, H^6 , $^3\text{J} = ^3\text{J}' = 7.6$), 8.17 (d, 1H, H^7 , $^3\text{J} = 7.6$), 8.31 (d, 1H, H^5 , $^3\text{J}' = 7.6$).

Compounds **19–22** were synthesized according this method.

2-Morpholin-4-yl-5-nitro-1H-isoindole-1,3(2H)-dione (19). Recrystallized from ethanol. M.p. = 221°C, Yield = 72%. IR (KBr) (ν , cm^{-1}) 1724 and 1721 (C=Oimide), 1540 and 1346 (NO_2). ^1H NMR (250 MHz, CDCl_3) (δ , ppm) 3.42 (t, 4H, CH_2N , $^3\text{J} = 4.3$), 3.89 (t, 4H, CH_2O , $^3\text{J} = 4.3$), 8.06 (d, 1H, H^7 , $^3\text{J}' = 8.0$), 8.63 (dd, 1H, H^6 , $^3\text{J}' = 8.0$, $^4\text{J} = 2.0$), 8.67 (d, 1H, H^4 , $^4\text{J} = 2.0$).

2-Phenyl-4-nitro-1H-isoindole-1,3(2H)-dione (20). Recrystallized from ethanol. M.p. = 121°C, Yield = 91%. IR (KBr) (ν , cm^{-1}) 1776 and 1734 (C=Oimide), 1545 and 1352 (NO_2). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 7.48–7.62 (m, 5H, H_{arom}), 8.16 (dd, 1H, H^6 , $^3\text{J} = ^3\text{J}' = 7.6$), 8.30 (dd, 1H, H^7 , $^3\text{J} = 7.6$, $^4\text{J} = 0.9$), 8.38 (dd, 1H, H^5 , $^3\text{J}' = 7.6$, $^4\text{J} = 0.9$).

2-Phenyl-5-nitro-1H-isoindole-1,3(2H)-dione (21). Recrystallized from ethanol. M.p. = 189°C, Yield = 94%. IR (KBr) (ν , cm^{-1}) 1780 and 1719 (C=Oimide), 1542 and 1342 (NO_2). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 7.48–7.63 (m, 5H, H_{arom}), 8.26 (d, 1H, H^7 , $^3\text{J} = 8.3$), 8.63 (d, 1H, H^4 , $^4\text{J} = 1.8$), 8.73 (dd, 1H, H^6 , $^3\text{J} = 8.3$, $^4\text{J} = 1.8$).

2-Benzyl-4-nitro-1H-isoindole-1,3(2H)-dione (22). Recrystallized from ethanol. M.p. = 141°C, Yield = 87%. IR (KBr) (ν , cm^{-1}) 1778 and 1720 (C=Oimide), 1538 and 1331 (NO_2). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 4.81 (s, 2H, CH_2), 7.30–7.38 (m, 5H, H_{arom}), 8.10 (dd, 1H, H^6 , $^3\text{J} = ^3\text{J}' = 7.6$), 8.22 (d, 1H, H^7 , $^3\text{J} = 7.6$), 8.32 (d, 1H, H^5 , $^3\text{J}' = 7.6$).

4-Amino-2-morpholin-4-yl-1H-isoindole-1,3(2H)-dione (23). To a solution of compound **18** (1.29 g, 4.65 mmol) in tetrahydrofuran (100 mL) was added catalytic quantity of 5% palladium on carbon. The reaction mixture was heated at 50°C and stirred under hydrogen atmosphere for 8 h. The catalyst was then filtered and the solvent evaporated under reduced pressure. The oily residue was purified by column chromatography over silica gel with dichloromethane and recrystallized from diethyl ether to give compound **23** as a yellow powder. M.p. = 264°C, Yield = 89%. IR (KBr) (ν , cm^{-1}) 3399 and 1623 (NH_2), 1773 and 1705 (C=Oimide). ^1H NMR (250 MHz, $\text{DMSO}-d_6$) (δ , ppm) 3.30 (t, 4H, CH_2N , $^3\text{J} = 4.3$), 3.72 (t, 4H, CH_2O , $^3\text{J} = 4.3$), 6.50 (bs, 2H,

NH₂), 6.96 (dd, 1H, H⁵, ³J = 7.0, ⁴J = 0.6), 7.02 (dd, 1H, H⁷, ³J' = 8.5, ⁴J = 0.6), 7.47 (dd, 1H, H⁶, ³J = 7.6, ³J' = 8.5).

Compounds **24-27** were synthesized according to this method.

5-Amino-2-morpholin-4-yl-1H-isoindole-1,3(2H)-dione (24). Recrystallized from diethyl ether. M.p. = 249°C, Yield = 80%. IR (KBr) (ν , cm⁻¹) 3454, 3353 and 1617 (NH₂), 1763 and 1703 (C=Oimide). ¹H NMR (250 MHz, CDCl₃) (δ , ppm) 3.40 (t, 4H, CH₂N, ³J = 4.9), 3.87 (t, 4H, CH₂O, ³J = 4.9), 4.40 (bs, 2H, NH₂), 6.84 (dd, 1H, H⁶, ³J = 8.0, ⁴J = 2.1), 7.02 (d, 1H, H⁴, ⁴J = 2.1), 7.60 (d, 1H, H⁷, ³J = 8.0).

4-Amino-2-phenyl-1H-isoindole-1,3(2H)-dione (25). Recrystallized from diethyl ether. M.p. = 180°C, Yield = 80%. IR (KBr) (ν , cm⁻¹) 3472, 3351 and 1630 (NH₂), 1753 and 1704 (C=Oimide). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 6.60 (bs, 2H, NH₂), 7.06-7.12 (m, 2H, H⁵ and H⁷), 7.42-7.58 (m, 6H, H⁶ and H_{arom}).

5-Amino-2-phenyl-1H-isoindole-1,3(2H)-dione (26). Recrystallized from diethyl ether. M.p. = 207°C, Yield = 68%. IR (KBr) (ν , cm⁻¹) 3492, 3373 and 1629 (NH₂), 1769 and 1690 (C=Oimide). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 6.61 (bs, 2H, NH₂), 6.90 (d, 1H, H⁶, ³J = 8.0), 7.04 (s, 1H, H⁴), 7.30-7.54 (m, 5H, H_{arom}), 7.62 (d, 1H, H⁷, ³J = 8.0).

4-Amino-2-benzyl-1H-isoindole-1,3(2H)-dione (27). Recrystallized from diethyl ether. M.p. = 147°C, Yield = 78%. IR (KBr) (ν , cm⁻¹) 3477, 3355 and 1634 (NH₂), 1744 and 1690 (C=Oimide). ¹H NMR (250 MHz, DMSO-*d*₆) (δ , ppm) 4.73 (s, 2H, CH₂), 6.53 (bs, 2H, NH₂), 7.00-7.05 (m, 2H, H⁵ and H⁷), 7.20-7.40 (m, 5H, H_{arom}), 7.47 (dd, 1H, H⁶, ³J = ³J' = 7.9).

Pharmacology

Drugs. All compounds were solubilized in DMSO and further diluted in RPMI ("Roswell Park Memorial Institute") medium (Sigma, St Quentin Fallavier, France) complemented with 1% L-glutamine (Gibco, BRL, Paisley, Scotland) and 10% heat inactivated fetal calf serum (FCS) (Sigma) referred as complete medium. Cyclosporine A (CsA) (Tocris, Illkirch, France) was dissolved in absolute ethanol containing 2% Tween 80 and this solution was further diluted in complete RPMI medium.

Splenocytes proliferation. Splenocytes were isolated from two spleens of 8-week-old female C57BL/6 mice (CR Janvier, Laval, France). Spleens were aseptically harvested and homogenized in a Petri dish containing HBSS medium (Sigma). Splenocytes suspension was

then hemolysed by buffer containing 20 mM Tris-HCl and 140 mM NH₄Cl. Cells were washed twice with RPMI, subsequently suspended in complete RPMI medium and seeded at densities of 1.5 × 10⁵/well in U-bottom 96-well culture plates (Falcon). Cells were incubated with 1 μg/mL concanavalin A (Sigma) in the presence of the studied compounds (90 μM) or CsA (5 μM) and cultured at 37°C in 5% CO₂ in a final volume of 150 μL of complete RPMI medium supplemented with 50 μM mercaptoethanol. Cell proliferation was assessed in sextuplicate after 72 h of culture, by colorimetric detection. Briefly, cells were incubated with 12.5 μg/well of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) for 4 h at 37°C. Formazan products were solubilized by 100 μL of lysis buffer (dimethylformamide (1V), SDS 20% (2V), pH 4.7) and overnight incubation at 37°C. Cell growth was assessed using a MRX microplate reader (Dynex Technologies, Chantilly, USA) with the test wavelength at 570 nm and expressed as optical density (OD) values. The inhibition of splenocytes proliferation was expressed as inhibitory rate [(OD value of control untreated cells - OD value of treated cells)/OD value of control untreated cells group] × 100.

Statistics. All results were compared by ANOVA analysis followed by a Dunnett test when the ANOVA test gives a significant difference ($p < 0.05$) between the different groups.

Results and discussion

Access to the cyclic urea analogues was achieved by a two-step method including the preparation and characterization of intermediate ureas. We implemented a process previously described by Gabriel et al. [10] consisting in the addition of a primary amine on a 2-chloroethyl or a 3-chloropropyl isocyanate. The resulting chloroalkylureas **2-9**, **28-32** and **38-41** are then cyclised by a nucleophilic substitution reaction in alkaline conditions to give the corresponding cyclic ureic compounds **10-17**, **33-37** and **42-45** (Scheme 1, Figure 2). The choice of this method was justified by the fact that this process has generated very few failures in the series previously synthesized in our laboratory [11,12]. The amines

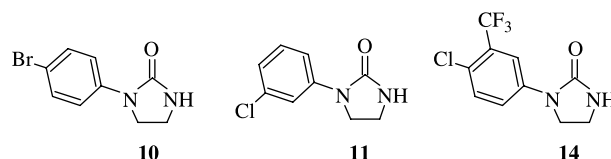
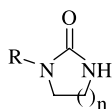


Figure 2. Structure of compounds 10, 11 and 14.

Table I. Inhibition of the mouse splenocyte ConA-induced proliferation by *N*-substituted imidazolidin-2-ones **10-17** and **33-37** and tetrahydropyrimidin-2(1*H*)-ones **42-45**



Compound (90 μ M)	R	N	Inhibition (%) \pm sem
10		1	60 \pm 1.8
11		1	43 \pm 1.9
12		1	< 30
13		1	< 30
14		1	91 \pm 2.1
15		1	< 30
16		1	< 30
17		1	< 30
33		1	ne
34		1	< 30
35		1	< 30
36		1	ne
37		1	< 30
42		2	< 30
43		2	< 30
44		2	< 30
45		2	< 30
	Compound 1 (90 μ M)		100 \pm 0.0
	Cyclosporine A (5 μ M)		100 \pm 0.0

ne: not evaluated

used were commercially available except for the aminophthalimides, which were prepared from the corresponding nitrophthalimides by catalytic reduction. Synthesis of the nitrophthalimides was accomplished by the action of an amine on a nitrophthalic anhydride in acetic acid [13,14,15].

The effect of drugs on mouse splenocytes proliferation was examined in order to determine the immunosuppressive potential with a rapid low-cost *in vitro* test. Freshly isolated murine splenocytes were stimulated with 1 μ g/mL ConA for 72 h in the presence of target cyclic ureas (90 μ M). Splenocytes were also treated with CsA (5 μ M) as a positive control. The results are shown in Table I. The molecules, which exhibited a lymphocyte proliferation inhibition percentage lower than 30%, have been considered inactive. Among the 17 tested compounds, three of them exerted a moderate (**10**, 60% and **11**, 43%) to potent (**14**, 91%) inhibitory activity. Generally speaking it seems that the molecules, which exert an immunosuppressive activity in this screening test show an imidazolidin-2-one scaffold *N*-substituted by a phenyl group. It seems however that the presence of a halogen on the phenyl moiety is favourable to the activity (**10**, **11** and **14**). These observations are in agreement with the data previously observed in imidazolidin-2-one series [9]. On the contrary the methylthio and the phenoxy group have demonstrated no interest (**12** and **13** respectively). Moreover the replacement of the phenyl substituent by a heterocyclic group triggered a loss of potency as it can be testified by compounds **15**, **16** and **17** and by phthalimidic derivatives **34**, **35** and **37**. Finally the elongation of the ureic cycle is responsible for a suppression of activity. Thus all the molecules of the tetrahydropyrimidin-2(1*H*)-one series (**42-45**) are inactive whereas some of their analogues in imidazolidin-2-one series were modestly to very potent [9] suggesting the importance of the imidazolidin-2-one scaffold in the induction of an immunosuppressive activity.

In conclusion, three new active molecules have been identified. The compound **14** exerts a potent activity in the ConA test with an inhibition of lymphocyte proliferation of 91%. Complementary studies are being investigated so as to confirm these results on human T lymphocytes. Other experiments must be realized in parallel on human MRC5 fibroblasts to determine the level of cytotoxicity of this molecule.

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